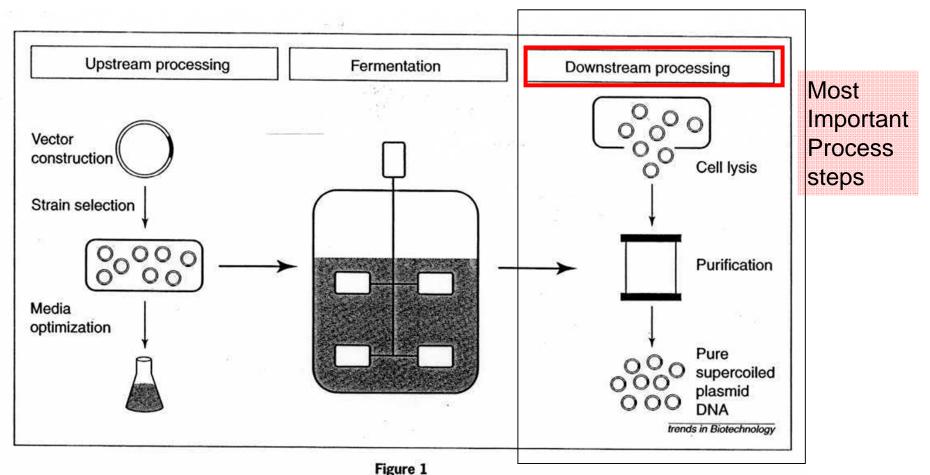
DNA as Therapeutic Agent

→ DNA Vaccines

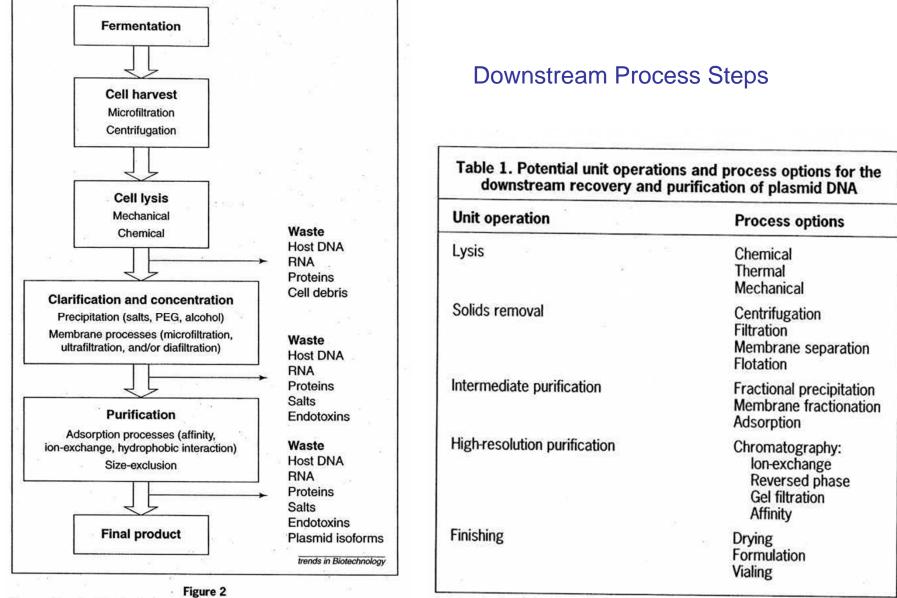
Application form: Plasmid DNA \rightarrow CCC-form

Main Challenge: ultra high purity; no contamination with non-desired genetic material



The three stages of plasmid-DNA process development.

Production of DNA



Process flow sheet for the large-scale purification of supercoiled plasmid DNA. Unit operations to be considered during process development are indicated together with the eliminated impurities.

Production of DNA

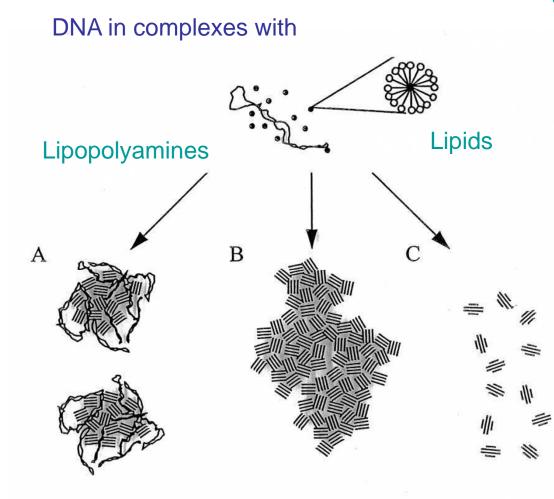
Purity requirements

Impurity	Recommended assay	Approval specification
Proteins	BCA protein assay	Undetectable
RNA	Agarose-gel electrophoresis	Undetectable
gDNA	Agarose-gel electrophoresis	Undetectable
	Southern blot	<0.01 µg (µg plasmid) ⁻¹
Endotoxins	LAL assay	<0.1 EU (µg plasmid) ⁻¹
Plasmid isoforms (linear, relaxed, denatured)	Agarose-gel electrophoresis	<5%
Biological activity and identity	Restriction endonucleases	Coherent fragments with the plasmid restriction map
	Agarose-gel electrophoresis	
	Transformation efficiency	Comparable with plasmid standards

Process step	Laboratory method	Large-scale process
Cell lysis	RNase, lysozyme	No enzymes Only GRAS reagents
Removal of cell debris	Centrifugation	Filtration, centrifugation or expanded bed chromatography
Removal of host impurities (RNA, gDNA, proteins and endotoxins)	RNase, proteinase K, organic solvents (phenol and chloroform)	Salting out PEG precipitation
Concentration	Alcohol precipitation	Alcohol precipitation, PEG precipitation
Plasmid purification	Ultracentrifugation (mutagenic reagents and ethidium bromide) IEC (gravity flow columns provided in commercial kits) RPC (organic, toxic solvents)	IEC and/or SEC (use only GRAS reagents)

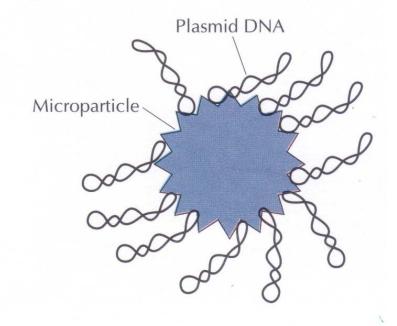
Improvement of DNA Transfer and Expression

Formulation of DNA



Charged polymeric microparticles

Figure **11.10** Schematic representation of the binding of plasmid DNA to the cationic surface of a polymeric microparticle.



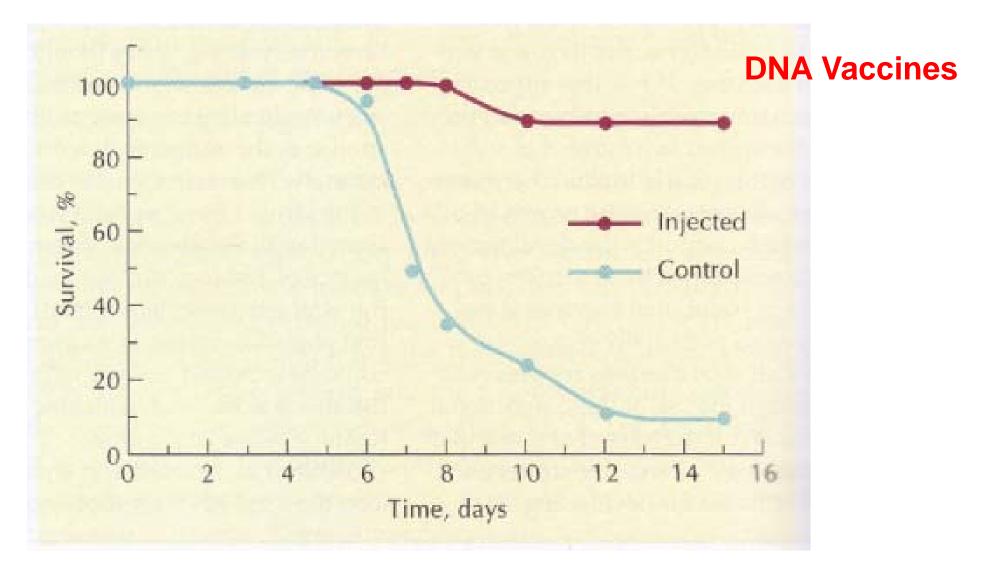


Figure 11.8 Survival of DNA-immunized mice. Injected mice were immuized with DNA that contained the influenza A virus nucleoprotein gene under the control of the Rous sarcoma virus promoter on an E. coli plasmid. The control mice were injected with vector plasmid DNA only. The x axis represents the number of days after animals were challenged with the live influenza virus.

Advantages of genetic immunization over conventional vaccines

- Cultivation of dangerous infectious agents is not required.
- Since genetic immunization does not utilize any viral or bacterial strains, there is no chance that an attenuated strain will revert to virulence.

• Since no organisms are used, attenuated organisms that may cause disease in young or immuno-compromised animals will not be a problem.

- Approach is independent of whether the microorganism is difficult to grow or attenuate.
- Production is inexpensive because protein does not need to be produced or purified.
- Storage is inexpensive because of the stability of DNA.
- One plasmid could encode several antigens/vaccines, or several plasmids could be mixed together and administered at the same time.

18.12.2014